

**TECHNICAL SHEET**

<b>B1147</b>		<b>BISMUTH SULPHITE AGAR</b>	
<b>Formula</b>			
<b>Ingredients:</b>		<b>gms/lit.</b>	
Peptic digest of animal tissue		5.00	
Pancreatic digest of casein		5.00	
Meat Extract B#		5.00	
Dextrose		5.00	
Disodium phosphate		4.00	
Ferrous Sulphate		0.30	
Bismuth Sulphite Indicator		8.00	
Brilliant Green		0.025	
Agar		20.00	
#- Equivalent to Beef extract			
Final pH (at 25°C): 7.6± 0.2			
<b>Directions:</b>			
Suspend 52.32 grams in 1000 ml purified/ distilled water. Heat to boiling to dissolve the medium completely. DO NOT OVERHEAT OR STERILIZE IN AUTOCLAVE or by fractional sterilization since overheating may destroy the selectivity of the medium. Transfer to a water bath maintained at about 50°C. The sensitivity of the medium depends largely upon uniform dispersion of precipitated bismuth sulphite in the final gel, which should be dispersed before pouring into the sterile Petri plates.			
<b>Principle:</b>			
In Bismuth Sulfite Agar, Meat Extract Band Peptic digest of animal tissue provide nitrogen, vitamins and minerals. Dextrose is an energy source. Disodium phosphate is a buffering agent. Bismuth sulfite indicator and brilliant green are complementary in inhibiting gram-positive bacteria and members of the coliform group, while allowing Salmonella to grow luxuriantly. Ferrous sulfate is for H <sub>2</sub> S production. When H <sub>2</sub> S is present, the iron in the formula is precipitated, giving positive cultures the characteristic brown to black colour with metallic sheen. Agar is a solidifying agent.			
<b>QC Tests – (I)Dehydrated Medium</b>			
	Colour:	Light yellow to greenish yellow	
	Appearance:	Homogeneous Free Flowing powder	
<b>(II)Rehydrated medium</b>			
	pH (post autoclaving/heating):	7.6 ± 0.2	
	Colour (post autoclaving/heating):	Yellow to greenish yellow	
	Clarity (post autoclaving/heating):	Opalescent gel with flocculent precipitate.	
<b>(III)Q.C. Test Microbiological</b>			
	Cultural characteristics observed after incubation at 30-35 °C for 24-48 hours. Recovery rate is considered as 100% for bacteria growth on Soyabean Casein Digest Agar.		
	MICROORGANISM (ATCC)	GROWTH	COLOUR OF COLONY
	Salmonella Typhimurium (14028)	Luxuriant	black or greenish-grey may have sheen
	Salmonella Abony (6017)	Good-luxuriant	Black with metallic sheen
	Salmonella enteritidis (13076)	Luxuriant	Black with metallic sheen
	Salmonella typhi (19430)	Luxuriant	Black with metallic sheen
	Enterobacter aerogenes (13048)	None - Poor	Brown to green*
	Escherichia coli (8739)	None - Poor	Brown to green*
	Shigella flexneri (12022)	None - Poor	Brown
	Enterococcus faecalis (29212)	Inhibited	---
	Key: * depends on inoculum density.		

Refer disclaimer Overleaf

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<b>Precautions :</b>	1. For Laboratory Use.				
	2. Follow proper, established laboratory procedures in handling and disposing of infectious materials.				
	3. HARMFUL. May cause sensitization by inhalation. Irritating to eyes, respiratory system and skin. Avoid contact with skin and eyes. Do not breathe dust. Wear suitable protective clothing. Keep container tightly closed.				
<b>Limitations :</b>	1. Since the nutritional requirements of organisms vary, some strains may be encountered that fail to grow or grow poorly on this medium.				
	2. It is important to streak for well isolated colonies. In heavy growth areas, <i>S. typhi</i> appears light green and may be misinterpreted as negative growth for <i>S. typhi</i> .				
	3. <i>S. typhi</i> and <i>S. arizonae</i> are the only enteric organisms to exhibit typical brown zones on the medium. Brown zones are not produced by other members of the Enterobacteriaceae. However, <i>S. arizonae</i> is usually inhibited.				
	4. Colonies on Bismuth Sulfite Agar may be contaminated with other viable organisms; therefore, isolated colonies should be subcultured to a less selective medium (e.g. MacConkey Agar).				
	5. Typical <i>S. typhi</i> colonies usually develop within 24 hours ; however, all plates should be incubated for a total of 48 hours to allow growth of all typhoid strains.				
	6. DO NOT AUTOCLAVE. Heating this medium for a period longer than necessary to just dissolve the ingredients destroys its selectivity.				
<b>Use :</b>	It is used for the selective isolation of Salmonellae from faeces, urine, sewage and other materials in accordance with United States Pharmacopoeia.				
<b>Storage :</b>	Dehydrated medium-below 30°C Prepared medium- Between 2 to 8°C. but not for more than two days as after which dye oxidizes to give green medium that could be inhibitory to some Salmonellae.				
<b>Packing :</b>	500 gm. bottle				
<b>Product profile:</b>	Reconstitution	Quantity on Preparation (500g)	pH (25°C)	Supplement	Sterilization
<b>B1147</b>	52.32g/l	9.556L	7.6± 0.2	NIL	DO NOT STERILIZE IN AUTOCLAVE

**Disclaimer:**

User must ensure suitability of the product(s) in their application prior to use. Products conform solely to the information contained in this and other related BIOMARKLABORATORIES publications.

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